Table S1. List of primers used in this study

Oligonucleotides	Description[a]
gehB 5′F	GGGGACAAGTTTGTACAAAAAAGCAGGCTAACA TAGGGCATAAGTGGAC
gehB 5′R	CGCTAACACTGACACCACG
gehB 3′F	GGTATCTGGCAAGTTAAACC
gehB 3′R	GGGGACCACTTTGTACAAGAAAGCTGGGTTTGCA CAACTCACTTCACC
Comp-*gehB*-F	TAGTGGTACCTGAGCGAATGGCTAGATGAA
Comp-*gehB*-R	GATCGAGCTCTGAACGCGTGAATAAAAACG
gehB-int-F	TCCAAATTATTGGGGTGGAA
gehB-int-R	ATGGTTGATTGGACGGATGT
gehB-noSP-F	TAGTCATATGTCGGAAAAAACATCAAC
gehB 3′R-EcoRI	TTTGAATTCCAGCACGATTTACATAGC
S412A	AGGTACATCTTGTAGGGCATGCTATGG
S412A_antisense	GAATTGTTTGACCACCCATAGCATGCC

[a] Restriction sites in sequences are underlined.

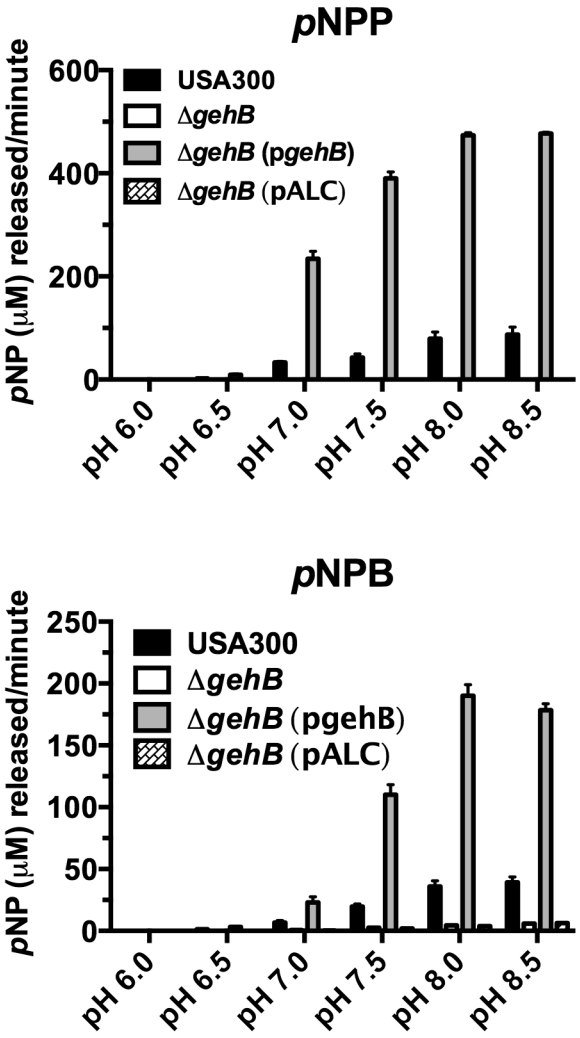

Figure S1: The optimal pH for SAL2 enzymatic activity ranges from pH 7.5 to pH 8.5. Lipase activity was assayed on culture supernatants from cells grown in TSB for 15 hours. Equivalents of OD_{600} 0.2 unit of culture supernatant were incubated with either (A) *para*-nitrophenyl palmitate (*p*NPP) or (B) *para*-nitrophenyl butyrate (*p*NPB) for 5-60 minutes at 37°C. Release of *para*-nitrophenol was measured by spectrophotometry at 410 nm and the lipase activity was calculated as the amount of *para*-nitrophenol released per minute. All enzymatic assays were measured on three biological and three technical replicates. The data are plotted as average ± the standard deviation. Δ*gehB* (pgehB) represents the USA300Δ*gehB* strain containing the complementing pgehB. Δ*gehB* (pALC) represents the USA300Δ*gehB* strain containing the empty pALC2073.